

PROXIMATE ANALYSIS AND DETERMINATION OF POLYCYCLIC AROMATIC HYDROCARBONS IN PROCESSED ANIMAL SKIN SOLD IN MAJOR MARKETS IN SOUTHWEST NIGERIA.

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ABSTRACT

This study aimed at determining the nutritional and level of chemical contaminants (heavy metals and Polyaromatic Hydrocarbons PAHs) uncommonly consumed animal skin sold in major markets in South-West, Nigeria. Proximate analysis (crude protein, moisture content, carbohydrate, ash content, oil content, nitrogen value and crude fibre) and heavy metals and PAHs concentrations were determined to evaluate the nutritional composition and safety of regular consumption of these products and to validate the notion that animal skin as a source of protein, can replace meat in meals. Samples of animal skin were purchased from three major markets in Lagos and Ogun State. Proximate composition was determined using standard analytical methods, while levels of heavy metal (Fe, Pb, Cd, Cr and As) and PAHs were using Atomic Absorption Spectroscopy and GC-MS respectively. The results of proximate composition were as follows (%); for the brown samples; the moisture content ($\leq 81.40 \pm 1.80$), ash content ($\leq 0.94 \pm 0.01$), oil content ($\leq 1.60 \pm 0.01$), crude protein ($\leq 3.75 \pm 0.85$), crude fibre ($\leq 1.25 \pm 0.01$) and carbohydrate ($\leq 14.51 \pm 1.20$). The concentrations of heavy metals detected in the samples were as follows (mg/kg); Fe $\leq 0.068 \pm 0.001$, Cd $\leq 0.006 \pm 0.001$, Pb $\leq 0.004 \pm 0.001$ and Cr $\leq 0.004 \pm 0.001$, all were below the maximum permissible limit set by WHO and USEPA while, Arsenic was below the detection limit of the instrument. Seventeen PAHs were detected in the samples and all were above the maximum permissible limits with high health risk. It is therefore advised that cow skin be discouraged and more awareness created so that people will know the dangers to health posed by this harmful practice.

Keywords: Animal skin, Polyaromatic Hydrocarbons, Heavy metals, proximate analysis, GC-MS

INTRODUCTION

Food safety and sustainability of life can be threatened by contaminants in foods and one of the major channels through which food is contaminated is food processing. Food processing can have negative impact on the food product, and this is a worldwide challenge [1]. Foods are processed in order to improve the taste, preservation, readily edible and remove unwanted substances. One of the food products that are vulnerable to contamination during

processing in Africa is animal skin. Shortage and high cost of meat has made people, especially in developing countries to find alternative in animal skin as replacement for meat [2]. In Nigeria, many believe that a meal is not complete without animal skin as it is included in major delicacy and in preparation of several stews especially at several public outings [3].

The processing of animal skin, also called singeing involves tenderizing the skin in hot water followed by shaving with razor blade or burning of the hair on the skin using firewood, tyres, plastics and spent engine oil mixed with firewood [2]. The burnt animal skin are scraped to remove ash and thereafter boiled in water for about one hour to obtain the finished product [4]. Contaminant may be incorporated into the animal skin during singing process, from the shavers used in removing the cow skin hair [5] and materials used in singeing [2, 3, 6, 7]. The residue of the chemical substances administer to diseased animal to treat ailment are retained in the hide even after slaughtering and processing [8].

Some of the toxic chemical contaminants that have been reported in processed animal skin are polyaromatic hydrocarbons (PAHs), dioxins, furan, and benzene [2, 6] and toxic metals especially lead [7]. Food processing of fats and proteins that involves smoking or high-temperature cooking is one of the major sources of PAHs contamination in foods. Burning of polystyrene plastics may release styrene vapour, which can readily be absorbed by the cow skin. Long-term exposure to styrene affects the central nervous system, causes headaches, depression, fatigue while short-term exposure can cause eye and throat irritation. Polychlorinated biphenyls (PCBs) and heterocyclic amines are also released when plastics are burnt, contaminating the cowhides [9, 10, 11].

Potential known human health effects associated with singeing of cowhide include; short-term tissue irritation (such as skin, respiratory, eyes and gastrointestinal), decreased fertility, developmental neurological effects and renal toxicity amongst butchers exposed to relatively high levels of PAHs [11]. The processors are also at risk because of the associated health hazards through inhalation of these toxic compounds. Significant levels of PAHs have been reported in processed foods [12]. Many of these PAHs (benzo[a]pyrene, benzo[a] anthracene, dibenzo[a,h]anthracene and chrysene) have been reported to possess carcinogenic and genotoxic properties [13, 14]. PAHs may result in biochemical disruptions of DNA and cell damage in the body [15] and can lead to mutations and development of tumors and cancerous cells [16, 17].

Health risk assessment of contaminants and residues in foods need to be monitored regularly. This is germane in order to ensure the safety of consumers in agreement with regulatory limits. Additionally, they can also help to reveal and determine new or unexpected emerging chemical substances [18].

The aim of this study is to determine the proximate composition, level of some heavy metals and Polyaromatic hydrocarbons (PAHs) in animal skin sold in major markets in Southwest, Nigeria. This is to ascertain the safety and nutritional value of this fondly eaten material. The findings of this study provide

information on the level of contaminants in cow skin and give answers to the pertinent question” How safe is the processed animalskin sold in the market?

MATERIALS AND METHODS

Sample Collection

Six samples of brown animal skin were purchased from different points of sale in three selected major markets in Ogun and Lagos States. This includes Ita ale market (Site A), central market (Site B) and Main Market (Site C). The purposive sampling method was adopted because the samples were sold at their points of sale in the market rather than at clusters. About 0.6kg of the animal skin was purchased from each seller and the samples were wrapped in aluminum foils, labeled, and transported to the laboratory in an ice-chest. The samples were kept in the freezer at 4°C before analysis.

Laboratory Analysis

Proximate Analysis

The analytical methods by Association of Official Analytical Chemists [19], for proximate analysis were adopted in this study. Moisture was determined using hot air oven at 105°C, until constant weight. Crude protein was determined using Kjeldahl method, total nitrogen was multiplied by a protein factor of 6.25. Dietary fiber was determined using the enzymatic gravimetric method. Ash was determined using gravimetric method by incinerating the samples at 550°C, until constant

weight. Total carbohydrate was determined according to the difference method by calculation

Heavy Metals Analysis of Samples

The concentrations of selected heavy metals (Lead (Pb), Cadmium (Cd), Copper (Cu), Iron (Fe) and Arsenic (As) in the animal skin samples were determined using Buck Scientific Atomic Absorption Spectrophotometer (model 210A) after acid digestion. One (1g) of each sample was digested with 10 mL of Aqua regia (1:3 HNO₃ and HCl) [19]. Standard solutions of the metals of interest were run in the instrument (AAS) for calibration before analysis.

Extraction of PAHs and Silica Gel Cleanup

The cow skin samples were allowed to dry and homogenized to powder, 5g of the homogenized sample was weighed and transferred into a beaker and 100ml of hexane: Acetone (1:1) was added. The sample was placed in an ultrasonic bath for 20 minutes and the mixture was allowed to settle and the solvent layer was decanted, the rotary evaporator was used to concentrate the mixture to 2ml and it was cleaned using a silica gel.

This method is based on EPA method 3630C. Silica gel was activated by heating at 130°C for 16 hrs. It was then stored in activated silica gel in a desiccators. A glass column was packed with 5g of silica gel and 1g of Anhydrous Na₂SO₄ was added. 20 ml of N-hexane was added to the column and eluted into

beaker. The 2ml sample extract was added to the top of the column quantitatively. Another 10ml of n-hexane was added to the column and eluted to waste. Before the column head dried out, 10 ml (1+1) Dichloromethane+Hexane was added and the eluent was collected. The eluent was then concentrated to 2 ml using a rotary evaporator and PAHs qualification and quantification was assessed using gas chromatography with mass spectrophotometry (GC-MS) [20].

Determination of PAHs

The identification and quantification of the PAHs in the extracts were carried out using gas chromatography (GC-MS Spectrometer model 6800) according to the method described by TNRCC (1997) available at the Nigerian Institute of Oceanography and Marine Research, Victoria Island, Lagos, Nigeria. The identification was based on the comparison of the retention times of the peaks with those obtained from the mixture of PAHs standards while; quantification was based on external calibration curves prepared from the standard samples of each of the PAHs.

Statistical Analysis

Data generated from proximate, heavy metal and polycyclic aromatic hydrocarbon analysis were evaluated using coefficient of variation statistical method, and the means were declared significantly different at 5% ($P \leq 0.05$) from [21].

RESULTS AND DISCUSSION

The result of proximate analysis is presented on Table 1 while concentration of heavy metals (Iron, Lead, Cadmium, Chromium and Arsenic) in the samples is presented in Table 2. Proximate analysis and the concentration of heavy metals were determined in the collected animal skin samples. This is to ascertain the concentration of these contaminants and their safety for consumption

Proximate Analysis

The results of proximate analysis of the collected animal skin samples from the three sites follow similar trend which is; moisture content>carbohydrate>crude protein>oil content>crude fibre>ash content. Moisture is sometime used for the estimation of the quality of food and it's one of the main factors for storage consideration due to the proliferation of microorganisms, such as fungi and mold. Total nitrogen consisted of protein nitrogen and little non-protein nitrogen. Ash content is the amount of total mineral residue left after incineration of samples until constant weight [22]. The study of moisture, protein, fat and ash contents of animal skin have a great significance for consumers and scientists, apart from knowing their nutritive value, also aids in understanding its better processing and preservation [23].

Table 1: Result of proximate analysis of the animal skin samples

Parameter	A	B	C
Moisture			
content @			
105°C	81.4 ±1.80	81.11 ±1.00	79.92 ±2.10
Ash content @			
450°C	0.94 ±0.01	0.82 ±0.01	0.81 ±0.01
Oil content	1.34 ±0.03	1.36 ±0.01	1.6 ±0.01
Crude Protein			
(%N*6.25)	3.75 ±0.85	3.75 ±0.02	1.68
Crude Fibre	1.23 ±0.01	1.25 ±0.01	1.18 ±0.01
Carbohydrate	10.74 ±0.05	11.11 ±0.03	14.51 ±1.20

The moisture and ash contents of all the collected samples from the three sites follows similar trend; C<B<A. Organisms that encourage food spoilage flourish well in foods with high moisture contents, thereby reducing the shelf life. Samples with high moisture content have a low shelf life because microorganisms can thrive and grow more in food with high moisture content [24, 25]. Ash content may reflect the amount of mineral or metallic composition in a sample [26]. Based on the fact that the concentrations of metallic content in the analyzed samples are low (Table 2), it can be inferred that the result of ash content of the samples is a reflection of the mineral content. High ash content of hides can be attributed to the

heat intensity generated from the processing methods. Diets with low fibre could cause constipation and eventually lead to disease of the colon like pile, appendicitis and cancer [27].

Oil content follow this trend; A<B<C, similar to carbohydrate content. Though, protein is detected in the animal skin samples, the amount of carbohydrate in the samples is higher than protein content. This is not in line with what has been reported by [28] but it's similar to the report of [29]. It is also observed that the animal skin samples have high carbohydrate content than protein, which contradicts what has been reported by [28, 29]. The protein content of the cow skin samples are low, this may be attributed to heat used in singeing the cow skin, resulting

in degradation of the essential amino acids of protein [30].

Result of Heavy Metals

Heavy metals determined in this study were detected in all the animal skin samples except As. The concentration of all the heavy metals was below the permissible limit of heavy metals in food as set by USEPA and W.H.O. The concentrations of Fe and Cr in the animal skin samples from sites A and C are similar, while Cd concentrations in the three sites follow this trend; A<B<C.

The result of Pb in this study is lower than [31] report with a value of 1.99 ± 0.86 ppm, but almost similar to the result of [29] with a value of 0.00-0.71 ppm while [5] had a very high value of 1.54 mgkg⁻¹. The results of Cd for this study is low

compared to the result of [29] (0.000-0.240 ppm), but when compared to WHO permissible limit, the level of Cadmium in the samples were found to be at a close range as the value which is (0.003) ppm and the USEPA maximum permissible limit of (0.005) ppm,

According to Ayanda et al. [31], the detection of Cadmium in animal skin could be as a result of the animal skin coming into contact with both soils and particles dispersed by the wind. Cadmium exposure is toxic to the kidney; it causes bone demineralization [33]. It also causes cancer of the bladder, pancreas, gall bladder and testes [33]. According to Okiei et al. [5] report, ingestion of Chromium may lead to vomiting, diarrhea, blood loss in the gastrointestinal tract, skin ulcers, eczema and respiratory lung cancer.

Table 2:Concentration of Heavy Metals in the Animal Skin samples

METALS	A	B	C	WHO	USEPA
Fe⁺²	0.068 ±0.001	0.064 ±0.001	0.067 ±0.001	0.01	0.3
Cd⁺²	0.001 ±0.003	0.002 ±0.003	0.006 ±0.001	0.003	0.05
Pb⁺²	0.002 ±0.002	0.001 ±0.002	0.004 ±0.002	0.01	0.015
Cr⁺²	0.004 ±0.006	0.002 ±0.006	0.004 ±0.006	0.05	0.1
As⁺²	BDL	BDL	BDL		
BDL– Below Detection Limit					

Result of the concentration of PAHs in the Animal Skin samples

Identification and quantification of PAHs in the collected animal skin samples was carried out to assess the presence of USEPA priority PAHs and evaluate the health risk associated with their consumption. The result is presented in Table 3.

Polycyclic aromatic hydrocarbons (PAHs) are a group of contaminants produced by burning of carbon-base materials. They can get into food either from the environment or during food processing. Some PAHs are known to cause cancer because they can damage DNA.

Seventeen types of PAH's compounds were detected in the animal skin samples from the three different markets, all exceeded the maximum permissible limits which is 12 ug/kg for carcinogenic PAH's. The level of Naphthalene, Dibenz[a,h]anthracene, and Dibenzo(a,h)pyrene are higher in A compared to B while C has the lowest. Level of Acenaphthylene, Chrysene, and Benzo[a]pyrene was found to be below detection level in all the samples. The level of Acenaphthene, Fluorene, Anthracene, and Benzo[c]phenanthrene are a little higher in C compared to A while B is the lowest. The level of Phenanthrene, 3-Methylcholanthrene, Benzo[ghi]perylene, and Dibenzo(a,i)pyrene was found to be higher in B compared to A while C is the lowest. The level of Fluoranthene, and Benzo[j+k+b]fluoranthene was found to be higher in A compared to C

while B is the lowest. The level of Pyrene, Benz[a]anthracene, Indeno[1,2,3-cd]pyrene, was found to be higher in B compared to C while A is the lowest.

The exposure limit for naphthalene as regulated by the National Institute for Occupational Safety and Health (NIOSH) is 10–15 µg/g. Hence, depending on the regularity of consumption and the amount consumed per time, irritation of the eyes, skin irritation, respiratory system discomfort, wheezing, shortness of breath, bronchitis, vomiting, kidney and liver damage could manifest as symptoms of systemic acenaphthylene overload in the consumers.

The results of this findings supports the work of Abdel-Shafy and Mansour, [34] which attributed the increase in concentration of PAHs in animal skin to accidental exposure during grazing of animal to contaminants in the grazing field. According to Adams et al. [15] the materials used in singeing may increase the concentration of PAHs in singed animal skin. PAH's can enter the body through inhalation, ingestion, and contact. Long term health effects of exposure to PAH's may include cataracts, kidney, and liver damage. Polyaromatic hydrocarbons(PAHs) also generates reactive intermediates such quinone, hydroxyl alkyl derivatives, which are not sufficiently polar to be excreted then tend to form covalent adducts with nucleic acid and lead to genetic effects.

Table 3: Levels of PAHs in the sampled animal skin ($\mu\text{g/kg}$)

PAH	A	B	C
*Naphthalene	40.0 \pm 0.5	20.1 \pm 0.005	16.0 \pm 0.005
*Acenaphthylene	BDL	BDL	BDL
*Acenaphthene	36.0 \pm 0.1	35.7 \pm 0.2	36.2 \pm 0.1
*Fluorene	16.1 \pm 0.1	15.8 \pm 0.2	16.2 \pm 0.1
*Phenanthrene	4.2 \pm 0.1	4.3 \pm 0.1	3.9 \pm 0.1
	32.2		
*Anthracene	\pm 0.001	31.8 \pm 0.2	32.4 \pm 0.1
*Fluoranthene	40.2 \pm 0.5	36.2 \pm 0.3	36.4 \pm 0.5
*Pyrene	20.2 \pm 0.3	20.5 \pm 0.4	20.3 \pm 0.2
Benzo[c]phenanthrene	32.3 \pm 0.4	32.1 \pm 0.5	32.6 \pm 0.3
**Benz[a]anthracene	56.2 \pm 0.5	60.1 \pm 0.4	56.4 \pm 0.5
**Chrysene	BDL	BDL	BDL
**Benzo[a]pyrene	BDL	BDL	BDL
Benzo[e]pyrene	55.8 \pm 0.6	56.2 \pm 0.5	56.4 \pm 0.3
**Benzo[j+k+b]fluoranthene	72.3 \pm 0.5	71.8 \pm 0.6	72.0 \pm 0.3
3-Methylcholanthrene	40.3 \pm 0.5	72.0 \pm 0.5	36.2 \pm 0.5
**Indeno[1,2,3-cd]pyrene	40.1 \pm 0.1	44.2 \pm 0.3	40.3 \pm 0.3
**Dibenz[a,h]anthracene	68.2 \pm 0.4	68.1 \pm 0.3	64.2 \pm 0.2
**Benzo[ghi]perylene	39.8 \pm 0.2	40.0 \pm 0.1	36.4 \pm 0.3
Dibenzo(a,h)pyrene	16.3 \pm 0.1	16.2 \pm 0.3	15.8 \pm 0.1
Dibenzo(a,i)pyrene	28.4 \pm 0.1	32.1 \pm 0.2	28.3 \pm 0.1
PAH4	128.5	131.9	128.4

Results of PAHs in this study are lower than the level reported by Okareh et al. [20] but, higher than the results of Oyekunle et al. [35] for PAHs in beef sausage.

Carcinogenic risk assessment of polycyclic aromatic hydrocarbons in animal skin

The cancer risk due to dietary exposure to PAHs was assessed using the individual PAH carcinogenic potencies(B(A)Pteq), the carcinogenic toxic equivalents (TEQ) and PAH4 index [35,36,37]. The Carcinogenic potencies of individual PAHs B(A)Pteqwas evaluated by multiplying the PAH concentration in the sample by the individual toxicity equivalency factor (TEF) (Eq. (3)). TheTEF is an estimate of the relative toxicity of individual PAH fraction compared to benzo(a)pyrene.

Carcinogenic potencies of individual PAHs (B(A)Pteq) = $C_i \times \text{TEF}_i$ (3)

PAH4 Index (PAH4) = $\frac{B[a]A + \text{Chr} + B[b]FL + B[a]P}{\text{B[a]A} + \text{Chr} + B[b]FL + B[a]P}$ (2)

The carcinogenic toxic equivalent (TEQs) was then obtained by summing the carcinogenic potencies of individual PAHs as indicated in Equation 4.

Carcinogenic toxic equivalents(TEQs) = $\sum(B(A)Pteq)$ (4)

The result of individual PAH carcinogenic potencies (B(A)Pteq) and carcinogenic toxic equivalents (TEQ) is presented in Table 4. These values are higher than the values reported by [35, 36].

The PAH4 index was assessed in this study based on the review by the Contaminants in the Food Chain (CONTAM) Panel, (2008). PAH4 is a more suitable indicator of PAHs in Food relating to occurrence and toxicity of PAHs in food,

Table 4: Results of Carcinogenic potencies (B(A)Pteq)and Carcinogenic Toxic Equivalent (TEQs) of the animal skin samples

PAHs	Conc.	B(A)Pteq						
		A	B	C	TEQ			
		(µg/kg)	(µg/kg)	(µg/kg)	A	B	C	(µg/kg)
Naphthalene	0.00	40.00	20.10	16.00	0.04	0.02	0.02	0.08
Acenaphthene	0.00	36.00	35.70	36.20	0.04	0.04	0.04	0.11
Fluorene	0.00	16.10	15.80	16.20	0.02	0.02	0.02	0.05
Phenanthrene	0.00	4.20	4.30	3.90	0.00	0.00	0.00	0.01
Anthracene	0.10	32.20	31.80	32.40	3.22	3.18	3.24	9.64

Fluoranthene	0.00	40.20	36.20	36.40	0.04	0.04	0.04	0.11
Pyrene	0.00	20.20	20.50	20.30	0.02	0.02	0.02	0.06
Benz[a]anthracene	0.10	56.20	60.10	56.40	5.62	6.01	5.64	17.27
Benzo[e]pyrene	0.10	55.80	56.20	56.40	5.58	5.62	5.64	16.84
Benzo[j+k+b]fluoranthene	0.01	72.30	71.80	72.00	0.72	0.72	0.72	2.16
Indeno[1,2,3-cd]pyrene	0.10	40.10	44.20	40.30	4.01	4.42	4.03	12.46
Dibenz[a,h]anthracene	5.00	68.20	68.10	64.20	341.00	340.50	321.00	1002.50
Benzo[ghi]perylene	0.01	39.80	40.00	36.40	0.40	0.40	0.36	1.16

The maximum level for the sum of PAH4 has been 12.00 µg/kg of a product (European Commission, 2002). The sum of PAH4 in samples A, B and C (µg/kg) is 128.5, 131.9 and 128.4 respectively. PAH4 in all the animal skin samples from the three sites are higher than the EFSA recommended maximum level.

CONCLUSION

This study determined the nutritional value of Cow skin, and the result of the proximate analysis shows that Cow skin samples analyzed had a very low protein content which is erroneously believe to be its major nutritional value. The results of this study reveal the presence of heavy metals in the animal skins though, below the maximum permissible limit of WHO and USEPA. Also 17 PAH's were detected in the cow skin samples and all were above the maximum permissible limits which

indicates their consumption should be discouraged because most of these Polyaromatic hydrocarbons are carcinogenic in nature and excessive consumption can lead to death. It is therefore advised that cow skin be discouraged, and more awareness created so that people will know the dangers to health posed by this harmful practice.

REFERENCES

- [1] Hussain, M. A. (2016). Food contamination: major challenges of the future.21 35
- [2] Ademola, A. S., Kayode, B. I., Motolani, A. M., and Muyideen, I. (2022). Consumers' Perception, Nutritional and Mineral Composition of Processed Cowhide (Ponmo) as Affected by Different Processing Methods. *Al-Qadisiyah Journal For Agriculture Sciences*, 12(1) pp. 65-69 <https://jouagr.qu.edu.iq/>
- [3] Acharya, A., Poudel, A., Sah, A. K., Maharjan, D., Tibrewal, S., Mandal, P. K., and Maharjan, D. (2016). Isolation and identification of bacteria from meat processing units of Kathmandu Valley. *International Journal of Microbiology and Allied Sciences*, 2016, 2.

- [4] Obayori, O. S., Salam, L. B., Oyetibo, G. O., Idowu, M., and Amund, O. O. (2017). Biodegradation potentials of polyaromatic hydrocarbon (pyrene and phenanthrene) by *Proteus mirabilis* isolated from an animal charcoal polluted site. *Biocatalysis and Agricultural Biotechnology*, 12, 78-84.
- [5] Okiei, W., Ogunlesi, M., Alabi, F., Osiughwu, B., and Sojinrin, A. (2009). Determination of toxic metal concentrations in flame treated meat products, ponmo. *African Journal of Biochemistry Research*, 3(10), 332-339.
- [6] Ofomata I.B., Nwankwo I.O., Ogugua A.J., Ezenduka E.V., Nwanta J.A., and Obidike R.I. (2020). Detection of polycyclic aromatic hydrocarbons in hide and skin of slaughtered cattle and goats in Anambra State, Nigeria. *Journal of Food Quality and Hazards Control*. 7:119-127.
- [7] Obiri-Danso. K, Hogorh N, and Antwi-Agyei.P, (2008). Assessment of contamination of singed hides from cattle and goats by heavy metals in Ghana. *AJEST* 2(8): 217-221.52
- [8] Tijani, S. A., and Ajayi, O. O. (2016). Perception of stakeholders to the proposed ban on cowhide consumption in Ogun State, Nigeria. *Journal of Agricultural Extension*, 20(1), 173-182.68
- [9] Antic, D., Jones, P. H., Michalopoulou, E., Papoula Pereira, R., Smith, R., Filli, K. B., and Rose, M. (2015). A study to review current evidence and outline work-streams to support the development of a policy for smoked, skin-on sheep meat
- [10] Hillard, S. (2016). *Why Are Americans Sick?*. Sylvester Hillard.34
- [11] Hill, T. D. (2015). Public Health Implications Associated with the Practice of Utilizing Tires to Singe Meat in Three Major Cities of Ghana: A Concurrent Mixed Methods Study.33
- [12] Hamidi, E. N., Hajeb, P., Selamat, J., and Razis, A. F. A. (2016). Polycyclic aromatic hydrocarbons (PAHs) and their bioaccessibility in meat: A tool for assessing human cancer risk. *Asian Pacific Journal of Cancer Prevention*, 17(1), 15-23.
- [13] Rengarajan, T., Rajendran, P., Nandakumar, N., Lokeshkumar, B., Rajendran, P., and Nishigaki, I. (2015). Exposure to polycyclic aromatic hydrocarbons with special focus on cancer. *Asian Pacific Journal of Tropical Biomedicine*, 5(3), 182-189.
- [14] Yebra-Pimentel, I., Fernández-González, R., Martínez-Carballo, E., and Simal-Gándara, J. (2015). A critical review about the health risk assessment of PAHs and their metabolites in foods. *Critical reviews in food science and nutrition*, 55(10), 1383-1405.
- [15] Adams I., Okyere D., and Teye M. (2013). Assessment of heavy metal residues in hides of goats singed with tyres, and the effect of boiling on the heavy metal concentrations in the hides.2
- [16] Appiah, A. I. (2016). Assessment of microbial quality and heavy metal levels of raw cattle hide and meat sold at retail outlets in Tarkwa, Western Region, Ghana. Unpublished MSc Thesis Department of Environmental Science, Kwame Nkrumah University of Science and Technology (KNUST), Kumasi, Ghana.
- [17] Nisha, A. R., Dinesh Kumar, V., Arivudainambi, S., Umer, M., and Khan, M. S. (2015). Polycyclic aromatic hydrocarbons in processed meats: a toxicological perspective. *Research Journal of Chemistry and Environment*, 19(6), 72-76.
- [18] Filazi, A., Yurdakok-Dikmen, B., Kuzukiran, O., and Sireli, U. T. (2017). Chemical contaminants in poultry meat and products. *PoultSci*, 15, 171.26
- [19] AOAC International. Official Methods of Analysis of AOAC International. 20th ed. Gaithersburg, MD, USA: AOAC International; 2016. p. 3172.
- [20] Okareh, O. T., Oshinloye, O. A., and Abiodun, D. (2021). Polycyclic Aromatic Hydrocarbon Accumulation in Meats Singed with Kerosene and Waste Tyres: A Case for

Public Health Concern in Nigeria. *World News of Natural Sciences*, 38, 49-59.

[21] Woko, E. C., Ibegbulem, C. O., and Alisi, C. S. (2020). Polycyclic Aromatic Hydrocarbon, Heavy Metal, and Derivable Metabolic Water and Energy of Cattle Hides Processed by Singeing.76

[22] Ganogpichayagrai A, and Suksaard C.(2020). Proximate composition, vitamin and mineral composition, antioxidant capacity, and anticancer activity of *Acanthopanax trifoliatum*. *J Adv Pharm Technol Res*. 2020 Oct-Dec;11(4):179-183. doi: 10.4103/japtr.JAPTR_61_20. Epub 2020 Oct 10. PMID: 33425701; PMCID: PMC7784940

[23] Mridha, M. A., Lipi, S. Y., Narejo, N. T., Uddin, M. S., Kabir, M. S., and Karim, M. (2005). Determination of biochemical composition of *Cirrhinusreba* (Hamilton, 1822) from Jessore, Bangladesh. *Journal of Science and Technology University Peshwar*, 29(1), 1–5.

[24] Adepoju, M. A., Omitoyin, B. O., Ajani, E. K., and Asha, K. (2018). Effect of smoking time and temperature on the proximate composition and quality of milkfish steaks. *Journal of aquatic food product technology*, 27(3), 369-378.4

[25] Okonkwo, S. I., and Opara, M. F. (2010).The Analysis of Bambara Nut (*Voandzeiasubterranea* (L.) thouars) for Sustainability in Africa.Africa Research Journal of Applied Sciences 5(6): 394 – 396

[26] Aletan, U. I., and Kwazo, H. A. (2019). Analysis of the proximate composition, anti-nutrients and mineral content of *Maeruacrassifolia* leaves. *Nigerian Journal of Basic and Applied Sciences*, 27(1), 89-96.

[27] Omosuli, S.V. (2014).Effects of processing on the chemical and Anti-nutritional properties of cassava roots. *Research and Reviews : Journal of Botanical Science* 3(2).27-31.59

[28] Akwetey W.Y, Eremong D.C, Donkoh E. (2013). Chemical and Nutrient Composition of Cattle Hide (“welle”) using different processing methods *J AnimSci Adv*. 3(4):176-180. 6

[29] Ijeoma, L., Ogbonna, P., and Effiong, I. D. (2015). Effects of various methods of singeing on the heavy metal, proximate and sensory properties of singed cow hide (Ponmo). *Scholars Research Library, Archives of Applied Science Research* 7(4): 44-50.

[30] Girard, J. P. (1992). *Technology of meat and meat products*. Ellis Horwood.28

[31] Ayanda, O., Ajayi, T., and Bilewu, O. (2021). Analysis of Some Heavy Metals in Cow Skin (Ponmo) Sold at Major Markets in Ado-Odo/Ota LGA, Ogun State, Nigeria: doi. org/10.26538/tjnpr/v5i6. 3. *Tropical Journal of Natural Product Research (TJNPR)*, 5(6), 1006-1009.

[32] Solidum, J.M., De Vera, M.J.D., Ar-Raquib, D.C. Abdulla, J.H.E. and Nerosa, M.J.V. (2013). Quantitative Analysis of Lead, Cadmium and Chromium found in Selected Fish marketed in Metro Manila, Philippines, *International J of Environmental Science and Development*; 4 (2).66

[33] Rodjana, C. (2016). Cadmium Exposure and Potential Health Risk from Foods in Contaminated Area. *Thailand. Toxicol. Res*, 32, 65-72.

[34] Abdel-Shafy, H. I., and Mansour, M. S. (2016). A review on polycyclic aromatic hydrocarbons: source, environmental impact, effect on human health and remediation. *Egyptian journal of petroleum*, 25(1), 107-123.

[35] Oyekunle, J. A. O., Yussuf, N. A., Durodola, S. S., Adekunle, A. S., Adenuga, A. A., Ayinuola, O., and Ogunfowokan, A. O. (2019). Determination of polycyclic aromatic hydrocarbons and potentially toxic metals in commonly consumed beef sausage roll products in Nigeria. *Heliyon*, 5(8), 02345.

[36] Kay, J. E., Cardona, B., Rudel, R. A., Vandenberg, L. N., Soto, A. M., Christiansen, S., and Fenton, S. E. (2022). Chemical effects on breast development, function, and cancer risk: Existing knowledge and new opportunities. *Current Environmental Health Reports*, 1-28.

[37] Tongo, I., Ogbeide, O., and Ezemonye, L. (2017). Human health risk assessment of polycyclic aromatic hydrocarbons (PAHs) in

smoked fish species from markets in Southern Nigeria. *Toxicology reports*, 4, 55-61.